

60.0 °C

## QuickGuide: RealFast<sup>™</sup> Variant Detection on AB QuantStudio 5

## Setup for Variant Detection RealFast<sup>™</sup> Assays:

- Open the QuantStudio Design & Analysis Software (QuickGuide is based on version 1.5.2) and click Create New Experiment.
- In Experiment Properties define Name: Name of experiment Instrument type: QuantStudio 5 System Block type: 96-Well 0.2 ml Block Experiment type: Standard Curve Chemistry: TaqMan<sup>®</sup> Reagents Run mode: Standard
- In Method select a sample volume of 20 µl and setup the PCR program according to the Instructions for Use (IFU) of your RealFast<sup>™</sup> Assay.
   Make sure Data Collection On is enabled.



- In Plate → Quick Setup → Plate Attributes define None as Passive Reference dye for Variant Detection RealFast<sup>™</sup> Assays (e.g. HLA-B27, HLA-B5701, other HLA-Assays).
- In Plate → Advanced Setup assign Targets and Samples to selected wells.
   > Select Targets and edit and enter assays. Keep default settings for reporter dyes: Target 1 = FAM (corresponds to FAM-labeled probe). Quencher = NFQ-MGB Target 2 = VIC (corresponds to HEX-labeled probe). Quencher = NFQ-MGB

» **Note**: In case you want to run several **different** RealFast<sup>TM</sup> Assays in parallel, add targets for each of the markers to be analyzed. For assignment of fluorophores to hydrolysis probes see Instructions for Use of the respective RealFast<sup>TM</sup> Assay. «

- Click Add repeatedly to enter all your samples and controls. Assign Targets and Samples and select the total number of wells by click+drag in the plate layout.
- Define your No Template Control (NTC). Select a replicate (2 wells) in the plate layout by click+drag. Select "N" for both targets in the Task field.
- Define your Samples and Controls (e.g. HLA-B27 Negative Control, HLA-B27 Positive Control). Select well(s) in the plate layout. Assign sample(s) to selected well(s). Select "U" for both targets in the Task field.

Quick Setup A			Advanced S	Setup						>		6	View	~
	Ta	argets			+ Add	Ø	Action	~				1	2	
		Name	Reporter	Quencher	Comments	Task	Quantity	'			A			
		HLA-B27	FAM	NFQ-MGB		~		3	×			Negativ	Negativ.	
		PCR Cont	VIC	NFQ-MGB		~			×		В	U	U	
											с	Positiv	Positiv.	1
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				Comments *1 +						D	Sample 1	Sample I	(	
	1	NTC						×		1		Complete	Complet	
		Positive	Control				×			E	U U			
		Negative Control									_			

- > Save the experiment.
- Load your PCR tubes / plate into the QuantStudio<sup>™</sup> instrument and press START RUN button to start the run.

## Analysis of Variant Detection RealFast<sup>™</sup> Assays:

After completing a run or after opening an **Existing Experiment** the software displays the **Results** tab.

- The Amplification Plot appears automatically.
- If you have **different** RealFast<sup>™</sup> Assays in one run, select the appropriate target by clicking **Action** and using the dropdown menu.
- Press the button Show Plot Settings that is indicated by an eye-symbol and adjust the parameters:
  - > Plot Type: ∆**Rn Vs. Cycle**
  - > Graph Type: Linear
     > Plot Color: Target
  - > Move to Analysis Settings.



- Disable Default Settings, Automatic Threshold and Automated Baseline. Select the individual targets and adjust the Threshold according to the IFU. Baseline Start Cycle should be set to "3" and Baseline End Cycle to "15". Adjust the threshold manually. Set the threshold value for the FAM channel just above the background fluorescent signal generated by the e.g. HLA-B27 Negative Control. Set the threshold value for the HEX channel at the onset of the exponential phase of the amplification curve.
- > Confirm the analysis settings by pressing the **Apply** button.
- > Review your samples by selecting individual wells in the plate layout.

» **Note:** Threshold settings of individual targets can be alternatively adjusted in the **Show Plot Settings** window. «

- To show results as table click the Well Table icon in the upper right corner. Adjust the table according to your needs by selecting/deselecting the listed features in the View dropdown menu.
- Verify absence of any contamination in the Negative Control. No amplification should be visible.

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#	Well	Sample Na	Target Na	Task	Ст	U
1	A1	NTC	HLAB27	NTC	Undeter	
1	A1	NTC	PCR Cont	NTC	Undeter	
2	A2	pos.Contr	HLAB27	UNKNOWN	20.815	
2	A2	pos.Contr	PCR Cont	UNKNOWN	21.401	

- To print a report, select **File > Print Report** in the upper menu bar:
  - > Select data for the report according to your needs.
  - > If you wish to export your data go to the **Export** tab and choose the file type and content